

CHEMICAL CONSTITUENTS OF THE NUDIBRANCH *CHROMODORIS MARISLAE*

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**Abstract:** The major metabolite of *Chromodoris marislae*, marislin (1a) was identified from spectral data and converted into the sponge metabolite pleraplysillin-2 (1b).

Nudibranchs are brightly-colored shell-less mollusks. Since they lack physical defences against predation, it has been assumed that the nudibranchs, like other opisthobranch mollusks,<sup>1</sup> employ a chemical defence mechanism.<sup>2</sup> The dorid nudibranch *Chromodoris marislae* Bertsch is colored white with bright orange spots.<sup>3</sup> The animals were collected by hand (-8 m) at various locations in the western Gulf of California.

The dichloromethane-soluble material (4.0 mg/animal) from a methanolic extract of *C. marislae* was chromatographed on Florisil to obtain marislin (1a, 2.0 mg/animal) as the major metabolite. More polar fractions were rechromatographed by LC on  $\mu$ -porasil to obtain four minor metabolites 2a,b and 3a,b, isolated as two inseparable pairs of isomeric esters.

Marislin (1a) was obtained as an optically-inactive oil having the molecular formula  $C_{20}H_{24}O_4$  (328.1690). The IR spectrum contained an ester band at  $1735\text{ cm}^{-1}$  and a strong olefinic band at  $1650\text{ cm}^{-1}$ . The  $^1\text{H}$  NMR spectrum (Table 1) contained signals that could be assigned<sup>4</sup> to the  $C_{15}$  acid portion of pleraplysillin-2 (1b).<sup>5</sup> By analogy to model compounds,<sup>6</sup> fifteen signals from the  $^{13}\text{C}$  NMR spectrum<sup>7</sup> of marislin (1a) were assigned to the acid portion of the molecule. The remaining five carbon signals at  $\delta$  96.3 (d), 105.2 (t), 105.6 (d), 152.7 (d) and 165.1 (s) were attributed to an acetal carbon, two carbons of an exocyclic methylene group and two carbons of an enol ether moiety. The  $^1\text{H}$  NMR signals for the  $C_5$  acetal portion of marislin (1a) were

assigned as the result of careful decoupling experiments. The band at 258 nm ( $\epsilon$  11,650) in the UV spectrum was attributed to the diene in the C<sub>5</sub> acetal portion of the molecule.

The structures of marislin (1a) and pleraplysillin-2 (1b) are formally related by a [3,3] sigmatropic rearrangement. This rearrangement occurred very slowly during routine handling but treatment of marislin (1a) with boron trifluoride etherate in anhydrous ether caused complete rearrangement to pleraplysillin-2 (1b). Pyrolysis of marislin at 290°C for 4 min. gave a 1:1:1 mixture of marislin (1a), pleraplysillin-2 (1b) and the acid (1, R = H). The 1:1 mixture of 1a and 1b could not be separated but the <sup>1</sup>H NMR spectrum served as an excellent model for those of the minor metabolite mixtures.

The mixture<sup>8</sup> of ketones (2a,b, 0.08 mg/animal) had the molecular formula C<sub>20</sub>H<sub>28</sub>O<sub>4</sub>. An IR band at 1725 cm<sup>-1</sup> was attributed to both ester and ketone groups. The <sup>1</sup>H NMR spectrum (Table 1) contained small (0.5 H) signals due to the 1:1 mixture of two C<sub>5</sub> units and larger signals assigned to a common C<sub>15</sub> acid portion. The signal at  $\delta$  0.89 (d, 6 H,  $J = 6$  Hz) coupled to a signal at 2.15 (m, 1 H) required a terminal isopropyl group, while the signal at 3.02 (s, 2 H) was assigned to the C-8 methylene protons. We therefore proposed the structure 2 for the C<sub>15</sub> acid portion of the ketone.

The mixture of lactones (3a,b, 0.06 mg/animal) had the molecular formula C<sub>21</sub>H<sub>26</sub>O<sub>6</sub>. The IR bands at 1790 and 1720 cm<sup>-1</sup> suggested the presence of the  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone and  $\alpha,\beta$ -unsaturated ester groups. The <sup>1</sup>H NMR spectrum (Table 1) contained a methoxy signal at  $\delta$  3.18 (s, 3 H), a signal at 6.68 (bs, 1 H) due to the  $\beta$ -proton on an  $\alpha,\beta$ -unsaturated lactone, coupled to a methyl signal at 1.93 (bs, 3 H), and an AB quartet at 2.54 (d, 1 H,  $J = 17$  Hz) and 2.61 (d, 1 H,  $J = 17$  Hz) due to the non-equivalent methylene protons at C-8. We proposed structure 3 for the C<sub>15</sub> portion of the lactone. The  $\gamma$ -methoxy- $\alpha,\beta$ -unsaturated  $\gamma$ -lactone functionality may be formed by air-oxidation of the corresponding furan in methanolic solution. Thus, the mixture of lactones 3a and 3b was probably an artifact of isolation.

*Chromodoris* species are known to eat sponges. The close structural relationship between marislin (1a) and pleraplysillin-2 (1b), a metabolite of the Mediterranean sponge *Pleraplysilla spinifera*, is therefore not surprising. It is remarkable, however, that the nudibranch should concentrate essentially only one metabolite, presumably a metabolite of an unidentified sponge.<sup>9</sup>

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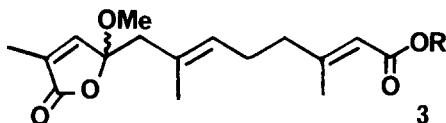
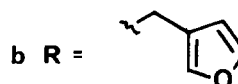
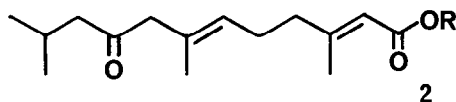
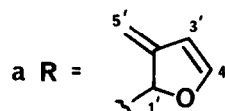
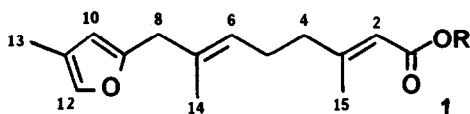


Table 1.  $^1\text{H}$  NMR Spectral Data (360 MHz,  $\text{CDCl}_3$ ,  $\delta$ ) for Marislin (1a), Pleraplysillin-2 (1b) and the Minor Metabolites 2 and 3.

H at C-#	<u>1a</u>	<u>1b</u>	<u>2a</u>	<u>2b</u>	<u>3a</u>	<u>3b</u>
2	5.72 (bs) <sup>a</sup>	5.69 (bs) <sup>b</sup>	5.73 (bs)	5.69 (bs)	5.73 (bs)	5.71 (bs)
4,5	≈ 2.2 (m, 4 H)		≈ 2.2 (m, 4 H)		≈ 2.2 (m, 4 H)	
6	5.18 (t, $J = 7$ Hz)		5.20 (t, $J = 7$ Hz)		5.17 (t, $J = 7$ Hz)	
8	3.22 (s, 2 H)		3.02 (s, 2 H)		2.54 (d, $J = 17$ Hz) 2.61 (d, $J = 17$ Hz)	
10	5.86 (s)		2.2 (m, 2 H)		6.68 (bs)	
11	-		2.15 (m)		-	
12	7.06 (s)		-		-	
13	1.98 (s, 3 H)		0.89 (d, 6 H, $J = 6$ Hz)		1.93 (bs, 3 H)	
14	1.60 (bs, 3 H)		1.61 (bs, 3 H)		1.65 (bs, 3 H)	
15	2.20 (bs, 3 H)		2.20 (bs, 3 H)		2.16 (bs, 3 H)	
1'	6.88 (s)		6.88 (s)		6.88 (s)	
3'	5.68 (s)		5.69 (s)		5.69 (s)	
4'	6.77 (s)		6.77 (s)		6.77 (s)	
5'	5.05 (s) 5.13 (s)		5.01 (s, 2 H) 5.03 (s) 5.15 (s)		5.01 (s, 2 H) 5.03 (s) 5.15 (s)	
-OMe	-		-		3.18 (s, 3 H)	

<sup>a</sup>All signals due to one proton unless otherwise designated. <sup>b</sup>See reference 4.

## REFERENCES AND NOTES

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2. M. R. Hagadone, B. J. Burreson, P. J. Scheuer, J. S. Finer, and J. Clardy, *Helv. Chim. Acta* 62, 2484 (1979).
3. H. Bertsch, A. J. Ferreira, W. M. Farmer and T. L. Hayes, *Veliger* 15, 287 (1973). Our sample was identified by J. R. Lance.
4. There is a systematic error between the  $^1\text{H}$  NMR chemical shift values reported for pleraplysillin-2 (1b) in  $\text{CCl}_4$  solution at 100 MHz<sup>5</sup> and those that we measured in  $\text{CDCl}_3$  solution at 360 MHz (Table 1).
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7.  $^{13}\text{C}$  NMR ( $\text{C}_6\text{D}_6$ )  $\delta$  188.8 (s, C-1), 165.1 (s, C-2'), 162.3 (s, C-3), 154.5 (s, C-9), 152.7 (d, C-4'), 138.2<sup>6</sup> (d, C-12), 133.0 (s, C-7), 125.4 (d, C-6), 120.7 (s, C-11), 115.4 (d, C-2), 109.2 (d, C-10), 105.6 (d, C-3'), 105.2 (t, C-5'), 96.3 (d, C-1'), 40.7 (t, C-4), 38.7 (t, C-8), 26.0 (t, C-5), 19.0 (q, C-15), 15.9 (q, C-14), 9.8 (q, C-13).
8. We did not attempt any heroic methods for separation of the mixtures due to lack of material (3-4 mg of each mixture). For the same reason, we were unable to record optical rotations or  $^{13}\text{C}$  NMR spectra for the minor metabolites 2 and 3.
9. We have not discovered a source of marisilin (1a) or pleraplysillin-2 (1b) among more than twenty common sponges of the Gulf of California.

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